Longer periods of charcoal incubation exerting deleterious effect on vegetative growth of rice

Somchai Butnan^{1,2}, Janista Duangpukdee^{1,2}, and Patma Vityakon^{2,3}

ABSTRACT: Charcoal application as a soil amendment in paddy does not improve rice growth and yield of the first crop, but it does in the following crops. Pre-incubation of charcoal prior to application to transplanted rice may enhance the synchronization of nutrient release and rice uptake. This study aimed at evaluating effect of charcoal incubation periods before rice transplanting on soil property and rice vegetative growth. A pot experiment was conducted under a greenhouse condition at the Field Research facility, Sakon Nakhon Rajabhat University during June - September 2019. Five pre-incubation periods of rice husk charcoal (RHC) before the rice transplanting were tested including no RHC with watering 0 day before rice transplanting and RHC with watering 0, 15, 30, and 60 days before rice transplanting (DBT). Soil pH and leaf number of rice were examined as preliminary data of the treatment effects on soil and plant. RHC amendment with watering 0 DBT had significantly higher soil pH and leaf numbers over other treatments. The longest the pre-incubation periods of RHC brought about the lowest rice leaf number. Nutrient deficient and volatile-matter-derived allelopathic effects were proposed as the mechanisms responsible for the deleterious effects of the RHC pre-incubation on the rice plants.

Keywords: Biochar, Nutrient deficiency, Oryza sativa, Soil pH, Synchronization

Introduction

Poor soil fertility produces low rice (Oryza sativa) yield in Northeast Thailand (Vityakon, 2007). Charcoal used as a soil amendment, namely biochar, has attracted much interest both in scientific and on-farm levels due to its soil fertility and crop yield improvement (Latawiec et al., 2017). Charcoal was proved to have agronomic benefits in soil properties (e.g., increases in soil pH and nutrient availability, and decreases in Al and Mn toxicities) and plant (e.g., tomato and corn) growth (Hossain et al., 2010; Butnan et al., 2015). Amongst many soil properties, a rise in soil pH is the most crucial role of charcoal, because pH is the master variable regulating all other soil properties (Bloom and Skyllberg, 2012).

Charcoal application to paddy rice did not improve yield in the first application, but it did in later seasons (Liu et al., 2016; Wang et al., 2018). Synchronization between nutrient release from charcoal and their uptake by rice plant could be the problem issue. Kongthod et al. (2015) demonstrated that the peak of NH_4^+ concentrations in a loamy-sand Nampong soil was at 60 days after additions of rice husk and cassava charcoal and that of NO3⁻ was at 90 days. These peaks of inorganic N release do not synchronize with nutrient requirement of rice, as shown by Murayama (1967) who reported that the maximum nutrient requirements of rice are in the tillering (35 - 40 days) and panicle formation (55 - 70 days) stages. Charcoal does not provide nutritional, especially N, benefit to first crop of rice. As a consequence, investment

¹ Plant Science Section, Faculty of Agricultural Technology, Sakon Nakhon Rajabhat University, Sakon Nakhon 47000. Thailand

² Soil Organic Matter Management Research Group, Khon Kaen University. Khon Kaen 40002, Thailand

³ Department of Soil Sciences and Environment, Faculty of Agriculture, Khon Kaen University, Khon Kaen 40002, Thailand

^{*} Corresponding author: Email: sbutnan@snru.ac.th

in charcoal amendment does not provide a pay off for farmers during the main rice-growing season (rainy season). Meanwhile, off-season rice incurs high production costs, for example average of 8,600 THB/rai in Ubon Ratchathani province (Srisurin et al., 2017).

This study therefore aimed at estimating the effect of periods of charcoal incubation before rice transplanting on soil pH and rice vegetative growth. We hypothesized that the more extended incubation periods prior to the transplanting would bring about higher rice growth.

Materials and methods

The soil used was Roi Et series collected from 0 - 15 cm-depth from a paddy field in Sakon Nakhon (17° 20' 36.7" N; 104° 02' 52.6" E). Charcoal was produced from rice husk under a kiln modified from the 200-liter metal cylindrical-shaped tank. A completely randomized design with three replications was conducted under a greenhouse in the Field Research facility of Plant Science section, Faculty of Agricultural Technology, Sakon Nakhon Rajabhat University. Data measurements were performed from June to September 2019. Treatments were composed of (i) no rice husk charcoal (RHC) amendment with watering 0 day before rice transplanting (No RHC + 0 DBT) and (ii) RHC amendment with watering 0 (RHC + 0 DBT), (iii) 15 (RHC + 15 DBT), (iv) 30 (RHC + 30 DBT), and (v) 60 (RHC + 60 DBT) days before the transplanting (Figure 1).

Each pot (height = 23.5 cm, top diameter = 19 cm, bottom diameter = 15 cm, v = 6,823 cm³ in a inverted truncated-pyramid shape) was filled with 6 kg air-dry soil and mixed thoroughly with RHC of 120 g/pot equal to 2% w/w according to Butnan et al. (2015). Prior to rice transplanting, each pot was watered to 70% of water holding capacity at the days of their respective treatments, except No RHC + 0 DBT and RHC + 0 DBT which were flooded to 3 cm-depth from the date of rice transplanting (**Figure 1**). Rice variety RD22 was seeded and nursed for 30 days in a nursery tray before transplanted to the pots.

Chemical fertilizers, graded 46-0-0, 0-0-60, and 18-46-0, were applied to all pots to achieve N, P, and K at the rates of 28.8 kg N/rai, 9.6 kg/ P_2O_5 /rai, and 14.4 kg K_2O /rai (Sun et al., 2015). Fertilizer application was divided into three periods – 50% of the total amount was at the rice planting, and the latter applied twice to 25% were at 20 and 40 days after the transplanting (Ly et al., 2015). Soil pH and rice leaf numbers were examined after rice transplanting till the end of the vegetative growth stage of the rice plant (72 DAP). Soil pH was measured using a portable pH meter. Sampling intervals of both parameters were at 44, 51, 65, and 72 days after rice planting.

Repeated measure analysis of variance based on the completely randomized design was used to evaluate the effects of periods of RHC incubation before rice transplanting on soil pH and leaf number of rice. Mean comparisons were performed using Fisher's least significant difference. Relationships between soil pH and



Figure 1 Timeline of rice growth stage and treatment managements; DAP = days after rice planting; DBT = days of pot watered before rice transplanting; RHC = rice husk charcoal.

Table 1 Repeated measure analysis of variance pertaining to the effects of incubation periods of rice
husk charcoal before rice transplanting (Treatment) and sampling intervals (Time) on soil
pH and leaf number per hill of rice over 72 days of rice planting.

	degree of freedom –	P-va	alue		
Source of variance		Soil pH	Leaf number/hill		
Treatment	4	**	***		
Time	3	ns	***		
Treatment x Time	12	*	***		
* = $P \le 0.05$; ** = $P \le 0.01$; *** = $P \le 0.001$; ns = not significantly different					

rice leaf numbers were assessed using Pearson's correlation. Significant differences were at $P \le 0.05$. The statistical analyses were performed by the SAS software version 9.1 (SAS Institute, Cary, NC, USA).

Results and discussion

Periods of RHC incubation before rice transplanting significantly affected soil pH and vegetative rice growth (leaf number per hill) (**Table 1**). Incubation of RHC at the transplanting date (RHC + 0 DBT) brought about significantly higher soil pH over other treatments at 72 DAP (**Figure 2a**). The highest leaf numbers in RHC + 0 DBT over other treatments were found from 44 to 72 DAP (**Figure 2b**). Soil pH significantly correlated to rice leaf number at 65 and 72 DAP (**Table 2**).

Charcoal constituted ash, volatile matter, and fixed carbon contribute to increases in soil pH (Butnan, 2015). Ash is composed of high content of alkalis, e.g., sylvite (KCl), calcite (CaCO₃), dolomite (CaMg(CO₃)₂), and magnesium carbonate (MgCO₂) (Yuan et al., 2011). Meanwhile, fixed carbon and volatile matter contain negative-charged functional groups which are attributed to high pH, e.g., carboxyl (-COO⁻), carbonyl (-CO⁻), and oxyl (-O⁻) (Yuan et al., 2011). In this study, higher soil pH of RHC + 0 DBT exerted increases in plant nutrient availability compared to treatments possessing longer periods of RHC incubation. Also, alkalis contain basic cations which are necessary to plant growth, e.g., K⁺, Ca²⁺, and Mg²⁺ (Butnan et al., 2015). Acid neutralization enriched the content of NH,⁺ which is the most favorite N form for rice uptake

44 DAP 51 DAP 65 DAP 72 DAP		Leaf number/hill				
		44 DAP	51 DAP	65 DAP	72 DAP	
Soil pH -0.151 ^{ns} 0.207 ^{ns} 0.444* 0.687**	Soil pH	-0.151 ^{ns}	0.207 ^{ns}	0.444*	0.687**	

 Table 2 Pearson correlations between soil pH and leaf number of rice in different time intervals after rice planting (n = 15).

* = $P \le 0.05$; ** = $P \le 0.01$; ns = not significantly different

DAP = days after rice planting

due to the enhancement of ammonification of charcoal-derived N and soil native N (Chen et al., 1998).

On the contrary to RHC + 0 DBT, the extended periods of RHC incubation prior rice transplanting, i.e., RHC + 15 DBT, RHC + 30 DBT, and RHC + 60 DBT, showed lower soil pH (Figure 2a) and rice leaf number (Figure 2b). Insufficient nutrients to support rice growth could result in the deleterious effects of the longer pre-incubation periods on the rice plants. Lower soil pH of the longer pre-incubation periods was a consequence of decreases in negative-charged organic anions in the decomposition process (Zimmerman, 2010; Dai et al., 2014). This acidified soil rendered

lower nutrient availability (Mengel and Kirkby, 2001). In addition, the lower rice growth may be driven by unavailability of ash-derived cations such as Ca²⁺ and Mg ²⁺. These basic cations could be hydrolyzed (Dai et al., 2014) and precipitated with RHC-derived silicon via the reaction: $Ca^{2+}/Mg^{2+} + SiO_4^{4-} = Ca/MgSiO_2 \cdot H_2O$ + H_oO (Morita et al., 2016; Antonangelo et al., 2017), prior the transplanting. Besides, N deficiency, which was triggered by N losses via ammonia volatilization and denitrification before having rice plants in the pots, may stunt rice growth after the transplanting. This was a consequence of neutralization under the preincubation treatments leading to acceleration of N mineralization and consequently N losses



Figure 2 Incubation periods of rice husk charcoal (RHC) treatments affecting soil pH (a) and leaf number of rice (b), i.e., no RHC with watering 0 day before rice transplanting (No RHC + 0 DBT), and RHC amendment with watering 0 (RHC + 0 DBT), 15 (RHC + 15 DBT), 30 (RHC + 30 DBT), and 60 (RHC + 60 DBT) days before rice transplanting. Vertical bars are the standard deviation. The accompanying tables beneath the graphic data demonstrate mean comparisons within time intervals (days after rice planting, DAP). The similar letters within a DAP are not significantly different (P≤0.05; Fisher's least significant differences).

(Dai et al., 2014).

In addition to pH effects, pre-incubation produce allelopathy treatments may to hasten the rice growth. These charcoal derived-allelopathic compounds included hydroquinones, benzoquinones, catechols, resorcinol, gallic acid, ethylene, acetylene, muconic acid, oxalic acid, maleic acid, malic acid, malonic acid, fumaric acid, formic acid, tartaric acid (Spokas et al., 2010; Olivier, 2011). Certain compounds were derived from both original pyrolysis and decomposition processes, e.g., quinones, catechols, ethylene, acetylene, gallic acid, and tartaric acid (Olivier, 2011; Kaal et al., 2012). Phenols, catechols, gallic acid, and quinones were reported to depress the rice growth (Djanaguiraman et al., 2005) via interfering various biochemical processes, examples, phosphorylation for pathway, ATPase activity, cell division, mineral uptake, and syntheses of carbohydrates, proteins, and nucleic acids (Einhellig, 2004).

Conclusions

The results of this study were opposite to our original hypothesis. The longer periods of incubation of rice husk charcoal before rice transplanting led to more deleterious effects on rice vegetative growth. Nutrient deficient and allelopathic effects were mechanisms detrimental to rice plants. The results clearly showed not to have pre-incubation of rice husk charcoal for paddy rice production. Data on the grain yield are necessary to validate the findings of this study.

References

- Antonangelo, J. A., J. Ferrari Neto, C. A. C. Crusciol, and L. R. F. Alleoni. 2017. Lime and calcium-magnesium silicate in the ionic speciation of an Oxisol. Sci. Agr. 74:317-333.
- Bloom, P. R., and U. Skyllberg. 2012. Soil pH and pH buffering. P. XIX1-14. In P. M. Huang, Y. Li and M. E. Sumner (eds). Handbook of Soil Sciences: Properties and Processes. CRC Press, Baca Raton.

- Butnan, S. 2015. Biochars differing in properties and rates impacting soil-plant and greenhouse gases in different textured and mineralogy soils. Ph.D. Thesis. Khon Kaen University, Khon Kaen.
- Butnan, S., J. L. Deenik, B. Toomsan, M. J. Antal, and P. Vityakon. 2015. Biochar characteristics and application rates affecting corn growth and properties of soils contrasting in texture and mineralogy. Geoderma 237–238:105-116.
- Chen, D. L., P. M. Chalk, J. R. Freney, and Q. X. Luo. 1998. Nitrogen transformations in a flooded soil in the presence and absence of rice plants: 1. Nitrification. Nutr. Cycl. Agroecosys. 51(3):259-267.
- Dai, Z., Y. Wang, N. Muhammad, X. Yu, K. Xiao, J. Meng, X. Liu, J. Xu, and P. C. Brookes. 2014. The effects and mechanisms of soil acidity changes, following incorporation of biochars in three soils differing in initial pH. Soil Sci. Soc. Am. J. 78(5):1606-1614.
- Djanaguiraman, M., R. Vaidyanathan, J. A. Sheeba, D. D. Devi, and U. Bangarusamy. 2005. Physiological responses of *Eucalyptus globulus* leaf leachate on seedling physiology of rice, sorghum and blackgram. Int. J. Agric. Biol. 7(1):35-38.
- Einhellig, F. A. 2004. Mode of allelochemical action of phenolic compounds. P. 217-238.
 In: F. A. Macías, J. C. G. Galindo, J. M. G. Molinillo, and H. G. Cutler (eds). Allelopathy: Chemistry and Mode of Action of Allelochemicals. CRC Press, Boca Raton.
- Hossain, M. K., V. Strezov, K. Y. Chan, and P. F. Nelson. 2010. Agronomic properties of wastewater sludge biochar and bioavailability of metals in production of cherry tomato (*Lycopersicon esculentum*). Chemosphere 78:1167-1171.
- Kaal, J., K. G. J. Nierop, P. Kraal, and C. M. Preston. 2012. A first step towards identification of tannin-derived black carbon: Conventional pyrolysis (Py–GC–MS) and thermally assisted hydrolysis and methylation (THM–GC–MS) of charred condensed tannins. Org. Geochem. 47:99-108.

- Kongthod, T., S. Thanachit, S. Anusontpornperm, and W. Wiriyakitnateekul. 2015. Effects of biochars and other organic soil amendments on plant nutrient availability in an Ustoxic Quartzipsamment. Pedosphere 25(5):790-798.
- Latawiec, A. E., J. B. Królczyk, M. Kuboń, K. Szwedziak, A. Drosik, E. Polańczyk, K. Grotkiewicz, and B. B. N. Strassburg. 2017. Willingness to adopt biochar in agriculture: The producer's perspective. Sustainability 9(655):10.3390/su9040655.
- Liu, Q., B. Liu, P. Ambus, Y. Zhang, V. Hansen, Z. Lin, D. Shen, G. Liu, Q. Bei, J. Zhu, X. Wang, J. Ma, X. Lin, Y. Yu, C. Zhu, and Z. Xie. 2016. Carbon footprint of rice production under biochar amendment – A case study in a Chinese rice cropping system. GCB Bioenergy 8(1):148-159.
- Ly, P., Q. Duong Vu, L. S. Jensen, A. Pandey, and A. de Neergaard. 2015. Effects of rice straw, biochar and mineral fertiliser on methane (CH₄) and nitrous oxide (N₂O) emissions from rice (*Oryza sativa* L.) grown in a rain-fed lowland rice soil of Cambodia: A pot experiment. Paddy Water Environ. 13(4):465-475.
- Mengel, K., and E. A. Kirkby. 2001. Principles of Plant Nutrition. Kluwer Academic Publishers, Dordrecht.
- Morita, M., W. Shinohara, R. Hashimoto, and S. Motoda. 2016. Microstructural analysis of initial scale formed on stainless steel sheet immersed in hot spring water. P. 9 17. In:
 S. Brankovic, J. Y. Kim, M. Shaoet al (eds). Electrodeposition for Energy Applications. The Electrochemical Society, New Jersey.
- Murayama, N. 1967. Nitrogen nutrition of rice plant. JARQ-JPN Agr. Res. Q. 2:1-5.

- Olivier, C. F. 2011. An investigation into the degradation of biochar and its interactions with plants and soil microbial community. M.S. Thesis. Stellenbosch University Stellenbosch, Stellenbosch.
- Spokas, K., J. Baker, and D. Reicosky. 2010. Ethylene: Potential key for biochar amendment impacts. Plant Soil 333(1):443-452.
- Srisurin, K., S. Boonkuson, and T. Kaykratok. 2017. Cost and return of off-season rice : A case study of Baan Hee, Moo.3, Kham Charoen sub-district, Trakan Phuet Phon district, Ubonratchathani province. P. 1201 - 1210. In: Ratchathani University National Conference, 26-27 July 2017. Ratchathani University, Ubon Ratchathani, Thailand.
- Sun, H., H. Zhang, D. Powlson, J. Min, and W. Shi. 2015. Rice production, nitrous oxide emission and ammonia volatilization as impacted by the nitrification inhibitor 2-chloro-6-(trichloromethyl)-pyridine. Field Crop Res. 173:1-7.
- Vityakon, P. 2007. Degradation and restoration of sandy soils under different agricultural land uses in Northeast Thailand: A review. Land Degrad. Dev. 18:567–577.
- Wang, C., J. Liu, J. Shen, D. Chen, Y. Li, B. Jiang, and J. Wu. 2018. Effects of biochar amendment on net greenhouse gas emissions and soil fertility in a double rice cropping system: A 4-year field experiment. Agric. Ecosyst. Environ. 262:83-96.
- Yuan, J.-H., R.-K. Xu, and H. Zhang. 2011. The forms of alkalis in the biochar produced from crop residues at different temperatures. Bioresour. Technol. 102:3488-3497.
- Zimmerman, A. R. 2010. Abiotic and microbial oxidation of laboratory-produced black carbon (biochar). Environ. Sci. Technol. 44(4):1295-1301.